TRADE SECRET

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STUDY TITLE: H-28548: Absorption, Distribution, Metabolism, and

Elimination in the Mouse

TEST GUIDELINES: U.S. EPA Health Effects Test Guidelines

OPPTS 870.7485 (1998)

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ORIGINAL REPORT

COMPLETED: November 3, 2010

REPORT REVISION 1

COMPLETED: April 21, 2011

PERFORMING LABORATORY: E.I. du Pont de Nemours and Company

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LABORATORY PROJECT ID: DuPont-18647-1017

WORK REQUEST NUMBER: 18647

SERVICE CODE NUMBER: 1017

SPONSOR: E.I. du Pont de Nemours and Company

Wilmington, Delaware 19898

U.S.A.

GOOD LABORATORY PRACTICE COMPLIANCE STATEMENT

This study was conducted in compliance with U.S. EPA TSCA (40 CFR part 792) Good Laboratory Practice Standards, which are compatible with current OECD Good Laboratory Practices, except for the item documented below. The item listed does not impact the validity of the study.

1. Qualitative analysis of urine samples for structure confirmation and elucidation was conducted on a non-GLP Liquid Chromatography/Mass Spectrometry (LC/MS) system. However, the identity of the parent analyte, the only analyte detected, was confirmed in urine samples using the test substance H-28548, which had a matching nominal mass-to-charge (m/z) ratio of approximately 329.

Sponsor: Study Director:	E.I. du Pont de Nemours and Company Wilmington, Delaware 19898 U.S.A. William J. Fasano, Sr., B.S. Senior Research Toxicologist	<u> 21-APR -2011</u> Date
Sponsor:	Sponsor Representative	Date
	Sponsor Representative	Date

QUALITY ASSURANCE STATEMENT

Work Request Number: 18647 Service Code Number: 1017

Key inspections for the above referenced study were completed by the Quality Assurance Unit of DuPont Haskell and the findings were submitted on the following dates:

Audit Dates	Date Reported to Study Director	Date Reported to Management
Protocol: March 17, 2010	March, 17, 2010	March, 17, 2010
Conduct: March 31, 2010 June 09, 2010	March 31, 2010 June 09, 2010	March 31, 2010 June 09, 2010
Report/Records: October 08, 11-13, 2010	October 13, 2010	October 14, 2010
Sponsor Edits 1: October 28, 2010	October 28, 2010	October 28, 2010
Report Revision 1: April 11, 2011	April 11, 2011	April 11, 2011

Reported by:

Antonio Pedulla

Quality Assurance Auditor

Date

CERTIFICATION

We, the undersigned, declare that this report provides an accurate evaluation of data obtained from this study.

LC/MS/MSDescription of the color o

LC/MS Metabolite ID by:

Timothy A. Snow, Ph.D.
Senior Research Chemist

2[-APR-200]
Date

Reviewed and Approved by:

Gary W. Jepson, Ph.D.

Manager

19-APR-2011

Date

Issued by Study Director:

William J. Fasano, Sr., B.S.
Senior Research Toxicologist

Date

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STUDY INFORMATION

Substance Tested: • HFPO Dimer Acid Ammonium Salt

• 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propionic

acid, ammonium salt

• 62037-80-3 (CAS Number)

• H-28548

Haskell Number: 28548

Composition: Proprietary

Purity: 84%

Physical Characteristics: Clear and colorless liquid

Stability: The test substance appeared to be stable under the

conditions of the study; no evidence of instability was

observed.

Study Initiated/Completed: March 16, 2010 / (see report cover page)

Experimental Start/Termination: March 16, 2010 / June 11, 2010

In-Life Initiated/Completed: March 31, 2010 / April 7, 2010

Notebook Number(s): E-114321-AM, E-114321-AI, E-98524-GE

REPORT REVISION 1

The elimination half-life $(T_{1/2})$ for H-28548 in male and female mice, following a single oral dose at 3 mg/kg, was estimated and reported.

SUMMARY

The absorption, distribution, metabolism, and elimination of H-28548 were investigated in the Crl:CD1(ICR) mouse. H-28548 was administered in water to 5 male and 5 female mice as a single oral dose at a target dose level of 3 mg H-28548/kg bodyweight (bw) and a dose volume of 10 mL/kg bw. Mice were housed individually in metabolism units and urine and feces were collected on dry ice predose and postdose at 0-6 hours, 6-12 hours, 12-24 hours, and every 24 hours until 168 hours post-dose. At 168 hours post-dose, mice were asphyxiated by exposure to carbon dioxide and then sacrificed by exsanguination. H-28548 was quantitated in urine, feces, and cagewash by liquid chromatograpy tamdem mass spectrometry (LC/MS/MS). Urine samples were further evaluted by LC/MS to confirm the identity of the parent analyte and determine if H-28548 was eliminated metabolized or unmetabolized.

Following oral administration of H-28548 in water, $30.8\% \pm 5.37\%$ and $39.3\% \pm 5.58\%$ of the administered dose was accounted for in urine (0-12 hours) from male and female mice, respectively. At the conclusion of the study (168 hours post-dose), the total accumulated amount of H-28548 detected in urine was $89.5\% \pm 6.91\%$ and $91.5\% \pm 6.04\%$ of the administered dose for male and female mice, respectively.

Elimination of H-28548 via urine accounted for a majority of the administered dose for both male and female mice; minor levels of H-28548 detected in feces from male $(2.00\% \pm 1.01\%)$ and female mice $(1.91\% \pm 0.85\%)$ were likely contamination from urine.

Cagewash, which is composed of dried excreta (urine and feces), accounted for $9.64\% \pm 3.99\%$ and $6.25\% \pm 3.16\%$ of the administered dose for male and female mice, respectively.

Following oral dosing with H-28548 in water and a 168 hour post-dose collection period, $101.2\% \pm 3.22\%$ and $99.7\% \pm 2.95\%$ of the administered dose was recovered from male and female mice, respectively.

Samples of urine evaluated using LC/MS were found to contain only the parent substance, H-28548. This finding, taken with recovery of the administered dose in urine, confirms that H-28548 was rapidly absorbed and eliminated unmetabolized following oral dosing in the mouse.

The elimination half-life $(T_{1/2})$ for H-28548 in male and female mice, following a single oral dose at 3 mg/kg, was estimated to be 21 and 18 hours, respectively.

INTRODUCTION

The data from this study provides basic information on the absorption, distribution, metabolism, and elimination (ADME) of H-28548 following oral dosing in the mouse.

OBJECTIVE

The objective of this study was to determine the ADME of H-28548 in the mouse following a single oral dose of H-28548 in water. Use of a non-radiolabeled test substance for determining a material balance and metabolite identification in the mouse is justified based on results from an *in vitro* metabolism experiment with rat hepatocytes and rat oral and rat and monkey intravenous dose kinetic studies, which suggests that H-28548 is not metabolized and is eliminated rapidly. (1,2,3,4)

ANIMAL WELFARE ACT COMPLIANCE

This study complied with all applicable sections of the Final Rules of the Animal Welfare Act regulations (9 CFR) and the Guidelines from the Guide for the Care and Use of Laboratory Animals (NRC 1996). All studies conducted by or for DuPont Haskell adhere to the following principles:

- The sponsor and/or the study director ensures that the study described in this report does not unnecessarily duplicate previous experiments, and is in compliance with the DuPont Policy on Animal Testing.
- Whenever possible, procedures used in this study have been designed to implement a
 reduction, replacement, and/or refinement in the use of animals in an effort to avoid or
 minimize discomfort, distress or pain to animals.
- DuPont Haskell policy is that animals experiencing severe pain or distress that cannot be relieved are painlessly euthanized, as deemed appropriate by the veterinary staff and study director or appropriate designee.
- Methods of euthanasia used during this study were in conformance with the above referenced regulation and the recommendations of the American Veterinary Medical Association (AVMA), 2007 Guidelines on Euthanasia.
- DuPont Haskell is accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care (AAALAC) International.

MATERIALS AND METHODS

A. Test Guidelines

The study design complied with the following test guideline:

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• U.S. EPA, OPPTS 870.7485. Metabolism and Pharmacokinetics, Health Effects Test Guidelines (1998)

B. Test Substance

The test substance (CAS registry number 62037-80-3) was supplied by the sponsor and assigned Haskell number 28548.

C. Test System

Male and female Crl:CD1(ICR) mice were obtained from Charles River Laboratories, Inc. (Raleigh, North Carolina, U.S.A.).

The Crl:CD1(ICR) mouse was chosen for this study because of the extensive experience with this strain and its suitability with respect to longevity, sensitivity, and low incidence of spontaneous diseases. Furthermore, the Crl:CD1(ICR) mouse has been used previously for toxicokinetic and toxicity testing of this chemical.

Each animal was assigned a unique identification number to be used throughout the study. The last 3 digits of the animal identification number were marked on the tail of each animal in indelible ink.

D. Animal Husbandry

1. Housing

During the pretest period, animals were housed individually in solid bottom caging with bedding. Animals were moved to metabolism units for the in-life phase of the study.

2. Environmental Conditions

Animal rooms were maintained at a temperature of 18-26°C (64-79°F) and a relative humidity of 30-70%. Animal rooms were artificially illuminated (fluorescent light) on an approximate 12 hour light/dark cycle.

3. Feed and Water

All animals were provided tap water *ad libitum* and fed PMI[®] Nutrition International, LLC Certified Rodent LabDiet[®] 5002 *ad libitum*. When housed in metabolism units, feed was supplied as ground chow.

4. Animal Health and Environmental Monitoring Program

As specified in the DuPont Haskell animal health and environmental monitoring program, the following procedures are performed periodically to ensure that contaminant levels are below those that would be expected to impact the scientific integrity of the study:

• Water samples are analyzed for total bacterial counts, and the presence of coliforms, lead, and other contaminants.

• Samples from freshly washed cages and cage racks are analyzed to ensure adequate sanitation by the cagewashers.

Certified animal feed is used, guaranteed by the manufacturer to meet specified nutritional requirements and not to exceed stated maximum concentrations of key contaminants, including specified heavy metals, aflatoxin, chlorinated hydrocarbons, and organophosphates. The presence of these contaminants below the maximum concentration stated by the manufacturer would not be expected to impact the integrity of the study.

The animal health and environmental monitoring program is administered by the attending laboratory animal veterinarian. Evaluation of these data did not indicate any conditions that affected the validity of the study.

E. Pretest Period

Upon arrival at DuPont Haskell, all mice were housed in quarantine. The mice were:

- quarantined for at least 6 days.
- identified temporarily by cage identification.
- weighed at least 3 times during quarantine and once prior to dosing.
- observed with respect to weight gain and any gross signs of disease or injury.

The animals were released from quarantine by the laboratory animal veterinarian or designee based on body weights and clinical signs.

F. Assignment to Groups

Animals were selected for use on study based on adequate body weight gain and freedom from any clinical signs of disease or injury. The weight variation of selected animals by sex was less than 4% of the mean weight.

Each animal was assigned an animal number and a cage identification number. The animal number and cage identification number were both included on the cage label.

At study start, the animals were at least 8 weeks old.

G. Dose Preparation, Analysis, and Rates

The test substance was prepared for administration by oral gavage. This route was chosen because it is most commonly used for toxicity studies with H-28548.

H-28548 was weighed into a vial (approximately 35.6 mg) and mixed with deionized water (100 mL). The dose solution was prepared at a nominal concentration of 0.3 mg H-28548/mL (adjusted for purity, 84%), with a target dose level of 3 mg/kg body weight (bw) and a dose volume of 10 mL/kg bw. The dose level was chosen based on the results of the 28-day daily oral

dosing study in mice, where the no-observed-adverse-effect level (NOAEL) was 0.1 and 3 mg/kg/day for males and females, respectively.⁽⁵⁾

The dosing solution was prepared prior to the day of use and was stored refrigerated at 1-10°C prior to dosing.

H. In-Life Phase

1. Material Balance and Tissue Distribution

The conduct of this study was designed to comply with the Tier 1 requirements of U.S. EPA, OPPTS 870.7485 - Metabolism and Pharmacokinetics, Health Effects Test Guidelines (1998).

Mice were housed individually in metabolism units and fasted for approximately 3 hours prior to dosing. Food was returned approximately 2 hours post-dose.

Five male and 5 female mice were administered H-28548 at a nominal target of 3 mg H-28548/kg bw. Two male and 2 female mice were each administered dose vehicle (deionized water at 10 mL/kg bw) for collection of control excreta and tissue samples. Mice were returned to individual metabolism units following dosing.

Urine and feces were collected on dry ice predose and at 0-6 h, 6-12 h, 12-24 h, and every 24 hours until 168 hours post dose. Evidence supporting a lack of metabolism of H-28548 in rat hepatocytes and rat oral dose administration studies, precluded the necessity for a radiolabeled form of H-28548 and collection of expired air.

At the end of the experiment (168 hours post dose), mice were killed by CO₂ asphyxiation followed by exsanguination. The following tissues (Tier 1) were collected:

liver
fat
G.I. tract (and contents)
kidney
spleen
whole blood
residual carcass

After collection, these samples were stored at approximately \leq -10°C.

Over the course of the experiment, residual feed was collected into a single container and stored refrigerated at 1-10°C. Cages were rinsed with deionized water, which was collected into a single container. Cage wash was stored at room temperature and/or refrigerated at 1-10°C.

I. Quantitation of H-28548

1. Sample Receipt

The dose solution, urine, feces, and cage wash samples were received and stored at approximately -20°C by the analytical laboratory upon receipt and when not in use.

2. Sample Preparation Procedure (dose solution and urine samples)

The frozen samples were thawed to room temperature and mixed briefly before sampling. A pipette was used to transfer 25 μ L of sample into an empty HPLC vial, and the sample weight was recorded to the nearest 0.0001 gram. The pipette was then used to add 975 μ L of HPLC grade water, and mixed. The initial sample preparation dilution factor = 1/sample weight (g). Additional sample dilutions were performed with HPLC grade water to ensure that the sample peak area results were within the calibration curve limits. Quality control fortification samples were also prepared at low, mid and high levels in control urine, and prepared for analysis using the same procedure.

3. Sample Preparation Procedure (cage wash samples)

The frozen cage wash samples were thawed to room temperature and mixed briefly before sampling. A pipette was used to transfer 200 μ L of sample into an empty HPLC vial, and the sample weight was recorded to the nearest 0.0001 gram. The pipette was then used to add 800 μ L of HPLC grade water, and mixed. The initial sample preparation factor = 1/sample weight (g).

4. Sample Preparation Procedure (feces samples)

The frozen feces samples submitted in 15-mL conical polypropylene centrifuge tubes were thawed to room temperature. HPLC grade water was added to the 13-mL mark, and the weight of water added was recorded to the nearest 0.01 gram. Five ball bearings (5/32" diameter) were added to the sample tubes and sealed. The samples were homogenized using a Genogrinder for 5 minutes at 1400 strokes/minute (SPEX CertiPrep Genogrinder 2000, Metuchen, New Jersey U.S.A.). After homogenization, the samples were placed in a refrigerator for overnight extraction. After overnight extraction the samples were shaken to mix and centrifuged for 10 minutes at 4150 rpm at 20°C. Approximately 1.5 mL of supernatant was added to a 1.7 mL microcentrifuge tube and further centrifuged for 15 minutes at 14,000 rpm and 20 °C. A syringe filter (PALL Acrodisc - 25 mm with 0.2 μ m Nylon Membrane) was then used to filter approximately 1 mL supernatant into a HPLC vial for analysis. The preparation factor = (H₂O weight (g) + feces weight (g)) / feces weight (g). Additional sample dilutions were performed with pooled control feces extract to ensure that the sample peak area results were within the calibration curve limits. Quality control fortification samples were also prepared at low, mid and high levels using 2 grams of control feces, and prepared for analysis using the same procedure.

5. Stock Solutions and Calibration Standards

A stock solution of H-28548 was prepared in HPLC grade water. The stock solution was diluted with HPLC grade water to prepare calibration standards at 0, 2.50, 5.00, 12.5, 25.0, 62.5, 156,

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and 250 ng/mL levels. The calibration standards used for the urine, cage wash, and dose solution sample types were prepared in HPLC grade water. The calibration standards used for the feces samples were prepared using control feces extract to correct for matrix effects.

6. **Instrument and Conditions**

The prepared samples were analyzed by LC/MS/MS using the following conditions:

Method 1 Quantitation of H-28548 in urine, feces, and cagewash

HPLC Instrument: Agilent Model 1100

HPLC Parameters:

Column (Urine, dose

Zorbax SB-C8; 2.1x100 mm with 3.5 micron particle size

solution, and cage wash): Column (Feces)

Zorbax SB-C8; 2.1x30 mm with 3.5 micron particle size

Mobile Phase:

A: 0.15% acetic acid in HPLC grade water

B: 0.15% acetic acid in acetonitrile

35 °C Column Temperature:

Injection Volume: 5 μL urine, dose and cage wash samples

2 μL for feces samples

HPLC Gradient	Total Time	Flow Rate	A	В
(Urine, dose solution,	(min)	$(\mu L/min)$	(%)	(%)
and cage wash samples)	0.00	400	65.0	35.0
	5.00	400	65.0	35.0
HPLC Gradient	Total Time	Flow Rate	A	В
(Feces samples)	(min)	$(\mu L/min)$	(%)	(%)
	0.00	400	95.0	5.0
	2.00	400	95.0	5.0
	2.10	400	70.0	30.0
	4.50	400	50.0	50.0
	6.00	400	5.0	95.0
	9.00	400	5.0	95.0
	9.10	400	95.0	5.0
	11.0	400	95.0	5.0

MS Parameters:

Ion Source: Turbo Spray, Negative Ion

120°C Temperature (TEM): Dwell 250 msec Curtain Gas Flow (CUR): 10.0 GS1: 25

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GS2: 25 IonSpray (IS) Voltage: -4500 6.00 **CAD** EP -10.0

Ouadrupole Resolution: Ouad. 1: Unit

Quad. 3: Unit

MRM Settings Q1 Mass Q3 Mass DP CE **CXP** H-28548 329.0 285.00 -20.0 -6.0 -7.0

7. **Quantitation**

The samples, calibration standards, and fortification quality control plasma samples were analyzed by LC/MS/MS. The calibration standard curve was generated by regression analysis using the chromatographic peak areas of the calibration standard solutions. The peak areas for the study samples and fortification QC samples were compared to the calibration standard curve to determine the concentration of the analyte. Any samples with peak areas above the upper calibration standard were diluted to ensure that the peak areas were within the calibration curve.

J. **Identification of Metabolites**

Samples of urine were pooled across animals for a given time interval where the mean percent of the administered dose (by sex) was $\geq 5\%$ (males and females: 0-6, 6-12, 12-24, 24-48, and 48-72 hours; feces extract samples were not pooled since the total mean percent of dose for each collection interval (by sex) was <5% of the administered dose.

Samples of pooled urine (25 µL) were diluted to 500 µL with Nanopure water prior to analysis. Samples of the diluted urine (20 µL) were qualitatively screened by LC/HRMS for metabolites. Retention time and mass spectral confirmation of the parent was performed by spiking control urine with approximately 40 ppm (v/v) of the test material (H-28548) and analyzing the spiked sample using the identical method for the study samples (Method 2).

1. Liquid Chromatography/Mass Spectrometry (LC/MS)

Method 2 Oualitative LC/MS Confirmation and Structural Elucidation of

metabolites in urine

Agilent 1100 HPLC with column thermostat and binary pump, HPLC/MS System:

> autosampler, variable wavelength detector (S/N DE63058654 -Agilent Inc., Little Falls, Delaware, U.S.A.). Thermo-Fisher OrbiTrap FT-MS (S/N 1016B - Thermo-Fisher Scientific Inc., San Jose, California, U.S.A.). The associated computer is loaded

with Thermo-Fisher Xcaliber Software (v 2.0.7)

HPLC Conditions:

Column: Agilent Zorbax SB-C18 column (2.1 x 150 mm) 3.5 µm particle

size

25°C Column Temperature:

Solvent A: 0.10% Acetic Acid in HPLC grade water

Solvent B:	0.10% Acetic acid in acetonitrile			
Gradient:	Time	A	В	
	(min)	(%)	(%)	
	0.0	98.0	2.0	
	20.00	0.0	100.0	
	25.00	0.0	100.0	
	25.10	98.0	2.0	
	30.00	98.0	2.0	
Flow Rate:	0.30 mL/min	n		
Run Time:	30.00 min			

 $\begin{array}{lll} Flow \ Rate: & 0.30 \ mL/mir \\ Run \ Time: & 30.00 \ min \\ Injection \ Volume: & 20 \ \mu L \\ UV \ Wavelength: & 190-400 \ nm \end{array}$

MS Conditions:

Collision Energy:

Ionization Mode: Electrospray negative ion

Source Voltage: 3.6 kV
Capillary Temperature: 330°C
Tube Lens voltage: 140 V
Source Current: 100 µA

Data Acquisition Function: Full Scan = 120-1000 Da (Profile mode), Mass Resolution =

30,000

Daughter Scans (Da)

Identity Daughters Start Mass End Mass of

90

500

H-28548 329 25 V daughter ion scan only

Scan Time Full scan 0.95 sec/scan; Daughter ion scan 0.3 sec/scan

Collision Gas and Pressure: Argon at 0.000602 mbar

2. Data processing

All chromatograms were screened for differences (chromatographic peaks) in control versus H28548-dosed urine samples using IntelliExtractTM; v. 12.0.1 (ACD, Toronto, Ontario, Canada) control-sample comparison software.

STATISTICAL AND DATA ANALYSIS

Group data were represented as a mean \pm SD.

The elimination half-life ($T_{1/2}$; time in hours to elimination of $\geq 50\%$ of the administered dose) for H-28548 in male and female mice was estimated by interpolation of (mean) cumulative urinary excretion data from 0 to 168 hours using Origin v7.0220 (OriginLab Corporation, Northhampton, Massachusetts, USA).

RESULTS AND DISCUSSION

A. Quantitation of H-28548 by LC/MS/MS

(Tables 1-2, Figures 1-3)

1. Calibration Standard Curve

A calibration curve for H-28548 is shown in Figure 1. The curve was generated based on resulting peak areas of the H-28548 analyte using a quadratic equation, and 1/x weighing.

2. Limit of Detection and Limit of Quantitation

The limit of detection (LOD) and limit of quantitation (LOQ) were determined by comparing the peak-to-peak noise in chromatograms of control matrix versus the signal of the lowest level calibration standard. The initial LOD was calculated as 3 times the concentration equivalent of the mean noise level. The initial LOQ was based on the lowest calibration standard concentration, which had at least a 10x signal-to-noise ratio. For a sample preparation factor of 1x the initial urine and cage wash sample LOD was 0.1 ng/g and for feces the initial LOD was 0.08 ng/g. For a sample preparation factor of 1x the urine, cage wash, and feces matrices all have an initial LOQ of 2.5 ng/g. The final LOD and LOQ for each sample was determined by multiplying the initial values by the sample preparation factor.

Example LOD & LOQ Calculation: Urine sample from animal 001M, 120 hour time point

- 25 μ L aliquot sample weight (g) = 0.0294 g
- Sample Preparation Factor = 1 / 0.0294 = 34.0
- Final LOD for this sample = $0.1 \text{ ng/g} \times 34.0 = 3 \text{ ng/g}$ (reported to 1 significant digit)
- Final LOQ for this sample = $2.5 \text{ ng/g} \times 34.0 = 85.0 \text{ ng/g}$ (reported to 3 significant digits)

Example LOD & LOQ Calculation: Feces sample from animal 001M, 120 hour time point

- Water Extraction Weight = 9.89 g. Feces weight = 2.869 grams
- Sample Preparation Factor = (9.89(g) + 2.869(g)) / 2.869(g) = 4.45
- Final LOD for this sample = $0.08 \text{ ng/g} \times 4.45 = 0.4 \text{ ng/g}$ (reported to 1 significant digit)
- Final LOQ for this sample = $2.5 \text{ ng/g} \times 4.45 = 11.1 \text{ ng/g}$ (reported to 3 significant digits)

Example LOD & LOQ Calculation: Cage wash sample from animal 001M, 168 hour time point

- 200 μ L aliquot sample weight (g) = 0.2097 g
- Sample Preparation Factor = 1 / 0.2097 = 4.77

• Final LOQ for this sample = $2.5 \text{ ng/g} \times 4.77 = 11.9 \text{ ng/g}$ (reported to 3 significant digits)

None of the predose urine or feces samples had detectable levels of H-28548.

3. Chromatographic Results (urine, cage wash, and dose samples)

H-28548 eluted as a well-resolved peak with a retention time of approximately 2.4 minutes. An example chromatogram for the lowest calibration standard at 2.5 ng/mL is shown in Figure 2a. An example chromatogram of a urine control matrix sample is shown in Figure 2b (H-28548 was not detected). A low level fortification quality control (QC) sample is shown in Figure 2c, which was fortified at a level of 400 ng/g, and had a preparation factor of 40x. A 24-hour urine sample from animal 001M, which had a total dilution factor of 1480x is shown in Figure 2d. The final concentration for this sample was 14,800 ng/g.

4. Chromatographic Results (feces samples)

H-28548 eluted as a well-resolved peak with a retention time of approximately 5 minutes. An example chromatogram for the lowest calibration standard at 2.5 ng/mL is shown in Figure 3a. An example chromatogram of a feces control matrix sample is shown in Figure 3b (H-28548 was not detected). A low level fortification quality control (QC) sample is shown in Figure 3c, which was fortified at a level of 250 ng/g, and had a preparation factor of 6.14x. A 12 hour feces sample from animal 001M, which had a total dilution factor of 34.0x is shown in Figure 3d. The final concentration for this sample was 775 ng/g.

5. Fortification QC Sample Results

The average QC fortification results for the urine matrix are provided in Table 1. The average recoveries for the low level, mid level, and high level fortification standards ranged from 101-102%. The associated coefficient of variation (CV) was 1% for each level and demonstrates acceptable method performance.

The average QC fortification results for the feces matrix are provided in Table 2. The average recoveries for the low level, mid level, and high level fortification standards ranged from 88-100%. The associated CV ranged from 2-4% and demonstrates acceptable method performance.

B. Dose Formulation Concentration, Animal Body Weights, Dosing Information

(Table 3, Appendices A-B)

The concentration of H-28548 in the dose solution, as confirmed by LC/MS, was 0.29 mg H-28548/mL, which was >96% of the nominal target (0.3 mg H-28548/mL).

At study initiation (day of dosing), males weighed 27.0 g \pm 0.47 g and females weighed 24.5 g \pm 0.52 g; the calculated dose rate for male (2.91 \pm 0.03 mg/kg bw) and female mice (2.91 \pm 0.06 mg/kg bw) were within 3% of the nominal target (3 mg/kg bw).

C. Urine Data

(Table 4, Figure 4, Appendix C)

Following oral administration of H-28548 in water, $30.8\% \pm 5.37\%$ and $39.3\% \pm 5.58\%$ of the administered dose (0-12 hours) was accounted for in urine from male and female mice, respectively.

At the conclusion of the study (168 hours post-dose), the cumulative amount of H-28548 detected in urine was $89.5\% \pm 6.91\%$ and $91.5\% \pm 6.04\%$ for male and female mice, respectively.

Elimination of H-28548 via urine accounted for the administered dose for both male and female mice.

D. Feces Data

(Table 5, Figure 5, Appendix D)

Following oral administration of H-28548 in water, the cumulative amount of H-28548 detected in feces over the entire collection period (0-168 hours) was $2.00\% \pm 1.01\%$ and $1.91\% \pm 0.85\%$ for male and female mice, respectively.

The minor amount of H-28548 detected in feces was likely contamination from of urine. Given the high levels of H-28548 in urine, and the design of the urine/feces collection system of the metabolism units, feces likely became contaminated with small amounts of urine when contacting surfaces in transit to the feces collection vessel.

E. Material Balance

(Table 6, Figure 6, Appendices E-F)

Following oral dosing with H-28548 in water and a 168 hour post-dose collection period, $101.2\% \pm 3.22\%$ and $99.7\% \pm 2.95\%$ of the administered dose was recovered from male and female mice, respectively.

Of the total H-28548 recovered, the majority of administered dose was account for in urine from both males (89.5% \pm 6.91%) and females (91.5% \pm 6.04%); lesser amounts of H-28548 were accounted for in feces (male = 2.00% \pm 1.01%; female = 1.91% \pm 0.85%). Cagewash, which is composed of dried excreta (urine and feces) accounted for 9.64% \pm 3.99% and 6.25% \pm 3.16% of the administered dose for male and female mice, respectively.

The carcass and residual feed were not analyzed for H-28548 because analysis of urine, feces and cagewash accounted for the majority of administered dose with an overall recovery of 100% \pm 10%.

F. Metabolite Identification

(Figures 7-9)

H-28548 was detected in its anionic form by negative ESI mass spectrometry. A representative reconstructed chromatogram of ions characteristic of H-28548 (parent) for the 6 hour female dosed mouse urine sample and control urine fortified with the H-28548 test substance is shown in Figure 7.

The LC/MS mass spectrum of H-28548 in urine shows a significant amount of its proton bound dimer (m/z 658.943 Da) and sodium bound dimer (m/z 680.923 Da) (Figure 8); the dimer and the sodium dimer were created in the MS system and were not present in the sample itself. The molecular anion (m/z 328.968) was observed in both urine from a mouse dosed with H-28548 and the urine fortified with the test substance H-28548, but at a low intensity relative to the dimer adducts. These dimers are not to be confused with a covalent dimer, such as the HFPO acid dimer parent, but are charged dimers sometimes formed, in-source, as a result of the desolvation and ionization processes necessary to be observed by electrospray ionization mass spectrometry.

The daughter ion mass spectra of the parent ion 328.97 Da for urine from a mouse dosed with H-28548 and urine fortified with the H-28548 test substance shows the same 2 characteristic fragment ions at m/z 284.977, the loss of CO₂ and 169.989, [C₃F₇] (Figure 9).

Subsequent to collection of the LC/MS, all sample data were screened for suspected metabolites manually and automatically for unexpected metabolites using the IntelliExtractTM control-comparison data processing tool. In all cases, there was no evidence of metabolism observed in any of the samples by either method and only the anionic form of the residual parent, H-28548, was detected.

G. Elimination Half-Life $(T_{1/2})$

(Appendix G)

The elimination half-life ($T_{1/2}$) for H-28548 in male and female mice, following a single oral dose at 3 mg/kg, was estimated to be 21 and 18 hours, respectively.

CONCLUSIONS

Following oral administration of H-28548 in water, $30.8\% \pm 5.37\%$ and $39.3\% \pm 5.58\%$ of the administered dose was accounted for in urine (0-12 hours) from male and female mice, respectively. At the conclusion of the study (168 hours post-dose), the total accumulated amount of H-28548 detected in urine was $89.5\% \pm 6.91\%$ and $91.5\% \pm 6.04\%$ of the administered dose for male and female mice, respectively.

Elimination of H-28548 via urine accounted for a majority of the administered dose for both male and female mice; minor levels of H-28548 detected in feces from male $(2.00\% \pm 1.01\%)$ and female mice $(1.91\% \pm 0.85\%)$ were likely contamination from of urine.

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Cagewash, which is composed of dried excreta (urine and feces), accounted for $9.64\% \pm 3.99\%$ and $6.25\% \pm 3.16\%$ of the administered dose for male and female mice, respectively.

Following oral dosing with H-28548 in water and a 168 hour post-dose collection period, $101.2\% \pm 3.22\%$ and $99.7\% \pm 2.95\%$ of the administered dose was recovered from male and female mice, respectively.

Samples of urine evaluated using LC/MS were found to contain only the parent substance, H-28548. This finding, taken with recovery of the administered dose in urine, confirms that H-28548 was rapidly absorbed and eliminated unmetabolized following oral dosing in the mouse.

The elimination half-life ($T_{1/2}$) for H-28548 in male and female mice, following a single oral dose at 3 mg/kg, was estimated to be 21 and 18 hours, respectively.

RECORDS AND SAMPLE STORAGE

Specimens (if applicable), raw data, the protocol, amendments (if any), and the final report will be retained at DuPont Haskell, Newark, Delaware, Iron Mountain Records Management, Wilmington, Delaware, or Quality Associates Incorporated, Fulton, Maryland.

REFERENCES

- 1. DuPont Haskell (2007). In Vitro Rat Hepatocyte Screen. Unpublished report, DuPont-23460.
- 2. DuPont Haskell (2008). Repeated Dose Oral Toxicity 7-Day Gavage Study in Rats. Unpublished report, DuPont-24009.
- 3. DuPont Haskell (2007). Biopersistence and Pharmacokinetic Screen in Rats. Unpublished report, DuPont-24281.
- 4. DuPont Haskell (2009). Cross-Species Comparison of FRD-902 Plasma Pharmacokinetics in the Rat and Primate Following Intravenous Dosing. Unpublished report, DuPont-17751-1579 RV1.
- 5. DuPont-Haskell (2008). A 28-Day Oral (Gavage) Toxicity Study of H-28397 in Rats with a 28-Day Recovery. Unpublished report, DuPont-24447.

TABLES

TABLES

EXPLANATORY NOTES

ABBREVIATIONS:

CV - coefficient of variation

NA - not applicable QC - quality control SD - standard deviation

Table 1 Mouse urine sample fortification QC results for H-28548

Mouse Urine	Fortification	Average	
Fortification	Concentration	Recovery	CV
Sample	(ng/g)	(%)	(%)
Low	400	102	1
Mid	100,000	100	1
High	1,000,000	101	1

Table 2 Mouse feces sample fortification QC result for H-28548

Mouse Feces	Fortification	Average	
Fortification	Concentration	Recovery	CV
Sample	(ng/g)	(응)	(%)
Low	250	88	4
Mid	1250	95	4
High	50,000	100	2
-	·		

Table 3 Dosing information

	Males		Fema	les
	Mean	SD	Mean	SD
Subject weight (g)	27.0	0.47	24.5	0.52
Test substance received (mg)	0.079	0.001	0.071	0.002
Dose (mg/kg bw)	2.91	0.03	2.91	0.06

Table 4 Urine, cumulative percent of dose

Post-Dose Time Point	Mal	es	Fema	ales
(hours)	Mean	SD	Mean	SD
Pre-dose	NA	NA	NA	NA
6	14.1	5.19	17.2	4.41
12	30.8	5.37	39.3	5.58
24	54.9	6.26	61.4	5.99
48	72.7	8.10	77.9	5.58
72	80.0	7.22	84.3	6.64
96	84.1	7.12	87.7	6.67
120	86.5	7.16	89.5	6.55
144	88.2	7.14	90.7	6.22
168	89.5	6.91	91.5	6.04

Table 5 Feces, cumulative percent of dose

Post-Dose Time Point	Males		Fema	Females	
(hours)	Mean	SD	Mean	SD	
0	NA	NA	NA	NA	
6	0.31	0.4	NA	NA	
12	0.56	0.38	0.70	0.46	
24	0.86	0.37	0.89	0.55	
48	1.34	0.56	1.40	0.47	
72	1.54	0.63	1.63	0.58	
96	1.71	0.73	1.70	0.60	
120	1.80	0.78	1.80	0.71	
144	1.89	0.89	1.89	0.82	
168	2.00	1.01	1.91	0.85	

Table 6 Material balance, percent of dose

	Males		Females		
	Mean	SD	Mean	SD	
Urine	89.5	6.91	91.5	6.04	
Feces	2.00	1.01	1.91	0.85	
Cage Wash	9.64	3.99	6.25	3.16	
Total	101.2	3.22	99.7	2.95	

FIGURES

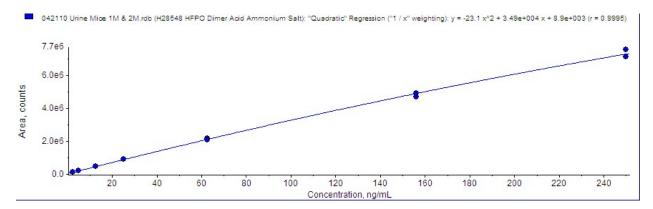
FIGURES

EXPLANATORY NOTES

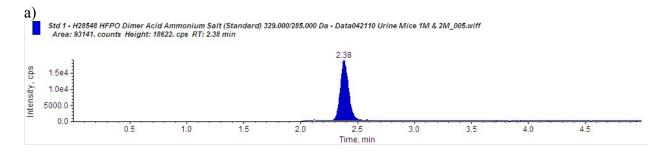
ABBREVIATIONS:

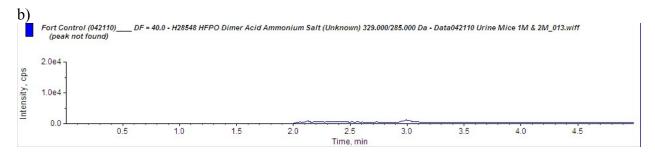
QC - quality control
cps - counts per second
m/z - mass-to-charge ratio
min - minute

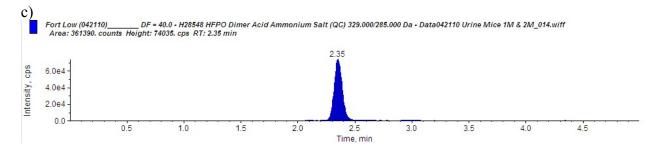
Figure 1 Calibration curve for H-28548



The LC/MS/MS chromatograms for a) lowest calibration standard at 2.5 ng/mL, b) urine control matrix sample, c) low level 400 ng/g fortification QC sample with preparation factor 40x, and d) a 24-hour urine study sample from animal 001M, which had a total dilution factor of 1480x and final concentration of 14,800 ng/g







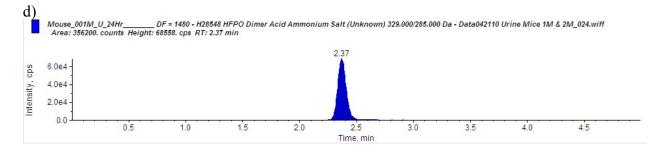
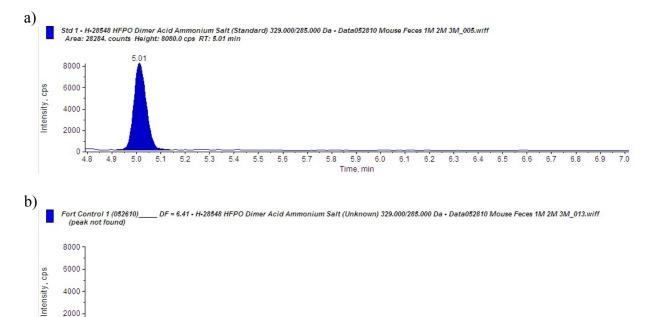
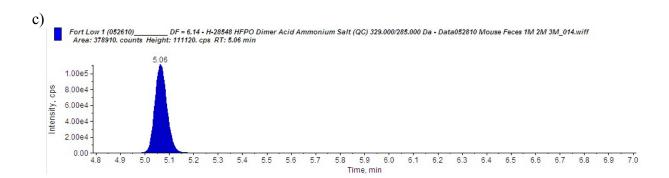


Figure 3

The LC/MS/MS chromatograms for a) lowest calibration standard at 2.5 ng/mL, b) feces control matrix sample, c) low level 250 ng/g fortification QC sample that had a preparation factor of 6.14x, and d) a 12-hour feces study sample from animal 001M, which had a 34x dilution factor and final concentration of 775 ng/g



4000 2000



5.9 6.0 Time, min

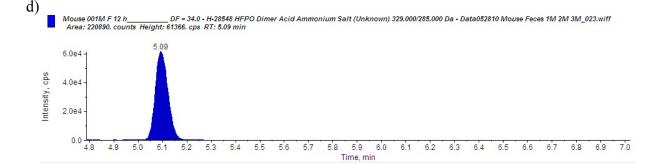


Figure 4 Urine, cumulative percent

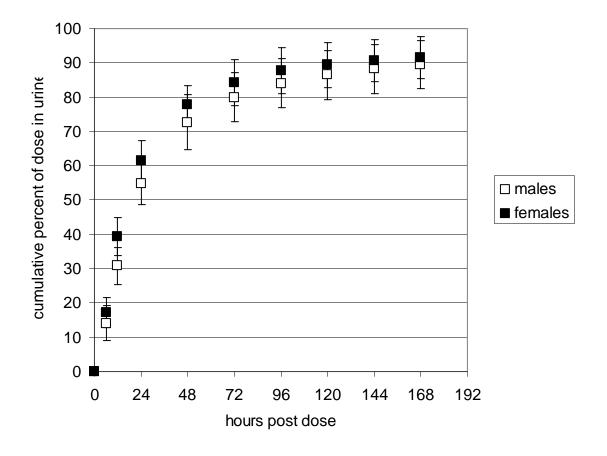


Figure 5 Feces, cumulative percent

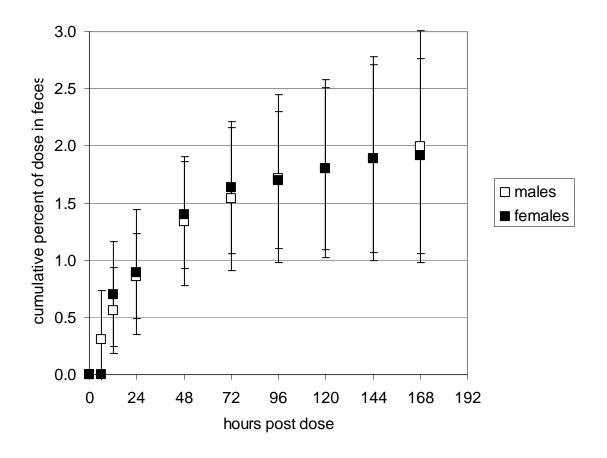


Figure 6 Material Balance, percent of dose

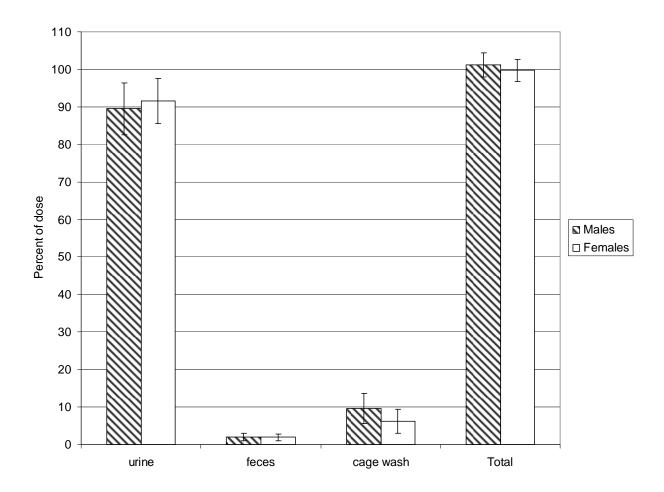


Figure 7
Reconstructed m/z 329 + 659 ion chromatograms characteristic of H-28548-dosed female mouse urine (6 hours after administration) – top and control mouse urine fortified with H-28458 test substance -bottom

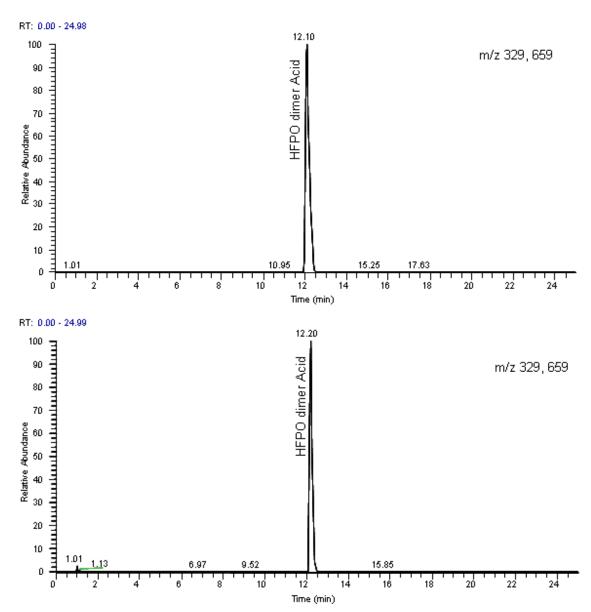


Figure 8
ESI negative mass spectra of H-28548 observed in dosed female mouse urine (6 hours after administration)—top; and control urine fortified with H-28548 test substance — bottom

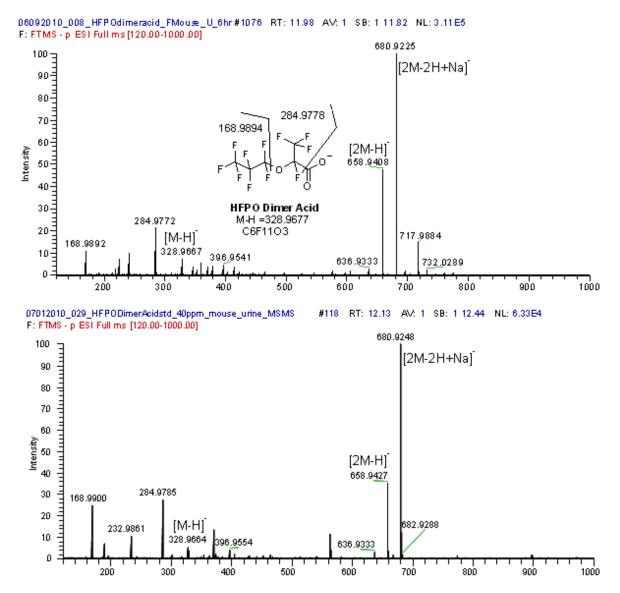
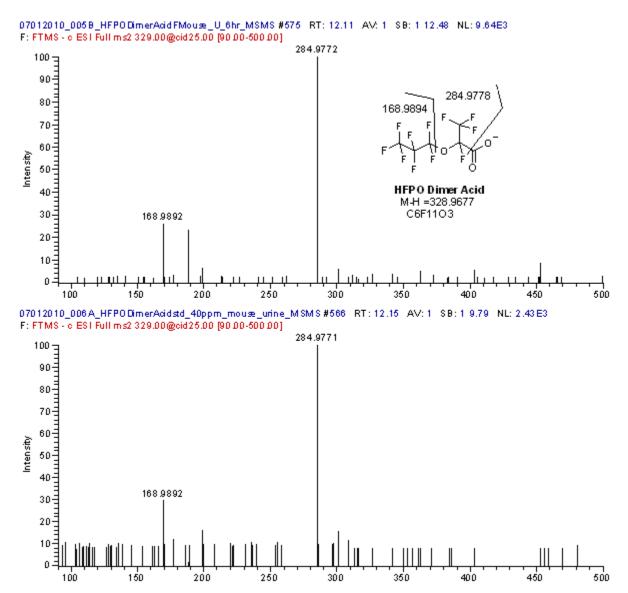


Figure 9
ESI negative daughter ion mass spectra of H28548 observed in dosed female mouse urine (6 hours after administration)—top; and control mouse urine fortified with H-28548 test substance — bottom



APPENDICES

APPENDICES

EXPLANATORY NOTES

ABBREVIATIONS:

F - female
h - hours

LOQ - limit of quantification
M - male
NA - not applicable
ND - not detected
SD - standard deviation

Appendix A Certificate of Analysis



E. I. du Pont de Nemours and Company Wilmington, DE 19898 USA

CERTIFICATE OF ANALYSIS

This Certificate of Analysis fulfills the requirement for characterization of a test substance prior to a study subject to GLP regulations. It documents the identity and content of the test substance. This work was conducted under EPA Good Laboratory Practice Standards (40 CFR 792).

Haskell Code Number H-28548

Common Name HFPO Dimer Acid Ammonium Salt

Purity Percent 84%

Other Components Water – 12.7%

Perfluorooctanoic acid – 150 ppm

Date of Analysis June 13, 2008

Expiration Date June 13, 2011

Instructions for storage NRT&H

Reference DuPont-25455

Analysis performed at E. I. DuPont de Nemours and Company

DuPont Haskell Laboratories

Newark, Delaware

USA

Approver;

Peter A. Bloxham, Ph.D.

Senior Research Chemist

Date

Revision #1: Revised COA expiration date based on compound stability assessment. 6/23/09

Appendix B Dosing Information

Dosing Information

Males Subject	Subject weight (g)	Compound received (mg)	Dose rate (mg/kg)
001M 002M 003M 004M 005M Mean	27.7 27.2 26.8 26.6 26.6 27.0	0.081 0.079 0.077 0.077 0.079 0.079	2.91 2.90 2.89 2.90 2.95 2.91 0.03
Females Subject	Subject weight (g)	Compound received (mg)	Dose rate (mg/kg)
001F 002F 003F 004F 005F Mean	24.3 24.2 24.5 24.1 25.4 24.5 0.52	0.073 0.070 0.070 0.071 0.073 0.071 0.002	3.01 2.88 2.85 2.94 2.89 2.91 0.06

Appendix C Urine Data

Urine Data - N	4ales
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Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
001M	80620	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.23 0.467 0.24 1.608 1.672 1.794 1.907 1.795 1.58	ND 36000 31300 14800 8060 2780 1720 1070 722 537	NA 16812 7512 23798 13476 4987 3280 1921 1141 966	NA 20.9 9.32 29.5 16.7 6.19 4.07 2.38 1.41 1.20 91.7	NA 20.9 30.2 59.7 76.4 82.6 86.7 89.0 90.5 91.7
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount	Percent	Cumulative (%)
002M	78880	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	2.155 0.599 0.311 1.403 2.12 2.489 2.833 2.866 2.595 3.139	ND 24300 32900 12000 6600 2720 1240 755 506 326	NA 14556 10232 16836 13992 6770 3513 2164 1313 1023	NA 18.5 13.0 21.3 17.7 8.58 4.45 2.74 1.66 1.30 89.2	NA 18.5 31.4 52.8 70.5 79.1 83.5 86.3 88.0 89.2
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
003M	77430	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.441 0.312 0.576 0.95 1.656 1.275 1.375 1.191 1.334 1.469	ND 29100 27300 18200 11500 3940 2460 1600 1020 669	NA 9079 15725 17290 19044 5024 3383 1906 1361 983	NA 11.7 20.3 22.3 24.6 6.49 4.37 2.46 1.76 1.27 95.3	NA 11.7 32.0 54.4 79.0 85.4 89.8 92.3 94.0 95.3

Urine	Data	_	Males

Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
004M	77140	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h	1.291 0.27 0.227 0.897 0.917 1.385 1.315 0.803 1.236 1.43	ND 26300 45800 19900 11500 4650 2470 2170 1140 924	NA 7101 10397 17850 10546 6440 3248 1743 1409	NA 9.2 13.5 23.1 13.7 8.35 4.21 2.26 1.83 1.71 77.9	NA 9.21 22.7 45.8 59.5 67.8 72.1 74.3 76.1 77.9
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
005м	78590	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.621 0.176 0.366 0.883 1.515 1.432 1.318 1.76 1.995 2.318	ND 46600 58500 21400 8560 3620 2100 1010 781 358	NA 8202 21411 18896 12968 5184 2768 1778 1558 830	NA 10.4 27.2 24.0 16.5 6.60 3.52 2.26 1.98 1.06 93.6	NA 10.4 37.7 61.7 78.2 84.8 88.3 90.6 92.6 93.6

Timenoint	Cumulative	
(hours)	Mean	SD
(110 01 0)	110411	- 55
0 h	NA	NA
6 h	14.1	5.2
12 h	30.8	5.37
24 h	54.9	6.26
48 h	72.7	8.10
72 h	80.0	7.22
96 h	84.1	7.12
120 h	86.5	7.16
144 h	88.2	7.14
168 h	89.5	6.91

Urine Data - Female	Urine	Data	-	Females	3
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Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
001F	73080	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.165 0.318 0.305 0.999 2.114 1.355 2.392 2.333 2.149 2.425	ND 28400 57500 19600 6260 2780 1230 486 270 183	NA 9031 17538 19580 13234 3767 2942 1134 580 444	NA 12.4 24.0 26.8 18.11 5.15 4.03 1.55 0.79 0.61 93.4	NA 12.4 36.4 63.1 81.3 86.4 90.4 92.0 92.8 93.4
	Total			Concentration			
Animal Number	H-28548 (ng)	Timepoint (hours)	Sample weight (g)	H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
002F	69600	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.164 0.414 0.29 0.958 1.856 1.442 1.931 2.51 2.853 1.959	ND 32500 45800 15600 6580 2490 1100 476 501 443	NA 13455 13282 14945 12212 3591 2124 1195 1429 868	NA 19.3 19.1 21.5 17.5 5.16 3.05 1.72 2.05 1.25 90.7	NA 19.3 38.4 59.9 77.4 82.6 85.6 87.4 89.4 90.7
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
003F	69890	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	2.575 0.287 0.41 0.873 1.975 2.041 2.368 3.804 2.232 1.971	ND 55400 45000 17300 4540 2580 736 334 231	NA 15900 18450 15103 8967 5266 1743 1271 516 345	NA 22.7 26.4 21.6 12.8 7.53 2.49 1.82 0.74 0.49 96.7	NA 22.7 49.1 70.8 83.6 91.1 93.6 95.4 96.2 96.7

Urine D	ata -	Females
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Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
004F	70760	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	0.706 0.481 0.251 0.642 1.447 1.384 1.433 1.553 1.718	ND 27200 50000 20800 6760 2490 1600 909 603 484	NA 13083 12550 13354 9782 3446 2293 1412 1036 736	NA 18.5 17.7 18.9 13.8 4.87 3.24 2.00 1.46 1.04 81.5	NA 18.5 36.2 55.1 68.9 73.8 77.0 79.0 80.5
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative
005F	73370	Pre-dose 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.152 0.266 0.483 1.092 2.702 4.6 5.534 3.029 2.981 3.908	ND 35900 35500 14600 5510 1490 548 396 254	NA 9549 17147 15943 14888 6854 3033 1199 757 660	NA 13.0 23.4 21.7 20.3 9.34 4.13 1.63 1.03 0.90 95.4	NA 13.0 36.4 58.1 78.4 87.7 91.9 93.5 94.5

Timepoint (hours)	Cumulative Mean	SD
0 h	NA	NA
6 h	17.2	4.4
12 h	39.3	5.58
24 h	61.4	5.99
48 h	77.9	5.58
72 h	84.3	6.64
96 h	87.7	6.67
120 h	89.5	6.55
144 h	90.7	6.22
168 h	91.5	6.04

Appendix D Feces Data

Feces	Data	_	Males

Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
001M	80620	0h 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.683 0.307 0.396 2.099 2.381 2.805 2.463 2.869 2.447 2.563	ND 512 775 95.1 54.7 28.9 13.5 <11.1 <13.0 <12.5	NA 157 307 200 130 81 33 NA NA	NA 0.19 0.38 0.25 0.16 0.10 0.04 NA NA NA	NA 0.19 0.58 0.82 0.98 1.09 1.13 1.13
Animal	Total H-28548	Timepoint	Sample	Concentration H-28548	Total Amount		Cumulative
Number	(ng)	(hours)	weight (g)	(ng/g)	(ng H-28548)	Percent	(%)
002M	78880	0h 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.478 0.367 0.701 1.694 3.18 3.207 3.302 3.173 2.993 3.291	ND 122 149 88.9 48.4 29.5 29.4 24.6 12.3 19.7	NA 45 104 151 154 95 97 78 37 65	NA 0.06 0.13 0.19 0.20 0.12 0.12 0.10 0.05 0.08 1.05	NA 0.06 0.19 0.38 0.58 0.70 0.82 0.92 0.96 1.05
	Total			Concentration			
Animal Number	H-28548 (ng)	Timepoint (hours)	Sample weight (g)	H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
003M	77430	0h 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.598 0.626 0.553 1.533 3.188 3.226 3.354 3.243 3.262 3.563	ND 1320 133 56.4 50.8 110 69.2 33.3 <9.88 <8.95	NA 826 74 86 162 355 232 108 NA	NA 1.07 0.09 0.11 0.21 0.46 0.30 0.14 NA NA 2.38	NA 1.07 1.16 1.27 1.48 1.94 2.24 2.38 2.38 2.38

reces Data - Maies	Feces	Data	_	Males
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Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
004M	77140	0h	1.716	ND	NA	NA	NA
		6 h	0.432	265	114	0.15	0.15
		12 h	0.945	122	115	0.15	0.30
		24 h	2.18	120	262	0.34	0.64
		48 h	3.803	275	1046	1.36	1.99
		72 h	3.745	43.5	163	0.21	2.20
		96 h	3.744	73.9	277	0.36	2.56
		120 h	3.734	30.7	115	0.15	2.71
		144 h	3.682	83.5	307	0.40	3.11
		168 h	4.069	76.6	312	0.40 3.51	3.51
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative
005M	78590	0h	1.971	ND	NA	NA	NA
		6 h	0.309	194	60	0.076	0.08
		12 h	0.713	554	395	0.50	0.58
		24 h	1.718	274	471	0.60	1.18
		48 h	3.088	126	389	0.50	1.67
		72 h	2.916	20.7	60	0.08	1.75
		96 h	2.943	19.6	58	0.073	1.82
		120 h	3.199	11.5	37	0.047	1.87
		144 h	3.159	<10.2	NA	NA	1.87
		168 h	3.358	11.1	37	0.047 1.92	1.92

Timepoint (hours)	Cumulative Mean	SD
0.1		
0 h	NA	NA
6 h	0.31	0.43
12 h	0.56	0.38
24 h	0.86	0.37
48 h	1.34	0.56
72 h	1.54	0.63
96 h	1.71	0.73
120 h	1.80	0.78
144 h	1.89	0.89
168 h	2.00	1.01

Feces	Data	_	Females

Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
001F	73808	0h 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.685 0.309 0.544 1.624 3.620 3.480 3.748 3.428 3.082 3.765	ND 663 97.4 75.6 99.2 19.7 27.9 <10.4 <10.5 <8.53	NA 205 53 123 359 69 105 NA NA	NA 0.28 0.07 0.17 0.49 0.09 0.14 NA NA NA	NA 0.28 0.35 0.52 1.01 1.11 1.25 1.25 1.25
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
002F	69600	0h 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h	1.714 0.494 0.421 1.454 3.517 3.279 3.457 3.576 3.767 3.093	ND 1640 107 152 37 90.5 17.9 58.8 48.1 13.6	NA 810 45 221 130 297 62 210 181 42	NA 1.16 0.06 0.32 0.19 0.43 0.09 0.30 0.26 0.06 2.87	NA 1.16 1.23 1.55 1.73 2.16 2.25 2.55 2.81 2.87
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
003F	69890	0h 6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h	1.635 0.303 0.5 1.178 2.807 3.128 3.01 3.452 2.569 3.105	ND 182 101 45.9 191 22.2 <10.5 10.9 <12.3 <10.1	NA 55 51 54 536 69 NA 38 NA	NA 0.08 0.07 0.08 0.77 0.10 NA 0.05 NA NA 1.15	NA 0.08 0.15 0.23 1.00 1.09 1.15 1.15

Feces	Data	_	Females

Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent	Cumulative (%)
004F	70760	0h	1.604	ND	NA	NA	NA
		6 h	0.5	592	296	0.42	0.42
		12 h	0.895	524	469	0.66	1.08
		24 h	1.283	134	172	0.24	1.32
		48 h	3.21	158	507	0.72	2.04
		72 h	3.42	57.5	197	0.28	2.32
		96 h	3.845	18.3	70	0.10	2.42
		120 h	3.698	29.7	110	0.16	2.57
		144 h	3.603	32.8	118	0.17	2.74
		168 h	3.898	11.7	46	0.06 2.80	2.80
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample weight (g)	Concentration H-28548 (ng/g)	Total Amount	Percent	Cumulative (%)
005F	73370	0h	2.015	ND	NA	NA	NA
		6 h	0.448	536	240	0.33	0.33
		12 h	1.051	256	269	0.37	0.69
		24 h	1.855	63.2	117	0.16	0.85
		48 h	3.817	67.5	258	0.35	1.20
		72 h	3.367	60.7	204	0.28	1.48
		96 h	3.699	<8.35	NA	NA	1.48
		120 h	3.217	<9.75	NA	NA	1.48
		144 h	3.495	<9.20	NA	NA	1.48
		168 h	3.346	<9.63	NA	NA 1.48	1.48

	0 1 1	
Timepoint	Cumulative	
(hours)	Mean	SD
0 h	NA	NA
6 h	NA	NA
12 h	0.70	0.46
24 h	0.89	0.55
48 h	1.40	0.47
72 h	1.63	0.58
96 h	1.70	0.60

120 h 144 h 168 h

Appendix E Cage Wash Data Cage Wash Data - 168 hours

Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample Weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent
001M 002M 003M 004M 005M	80620 78880 77430 77140 78590	168 h 168 h 168 h 168 h 168 h	274.506 274.514 214.319 207.445 214.205	32.3 30.8 24.4 55.7 17.4	8867 8455 5229 11555 3727 Mean SD	11.0 10.7 6.75 15.0 4.74 9.64 3.99
Animal Number	Total H-28548 (ng)	Timepoint (hours)	Sample Weight (g)	Concentration H-28548 (ng/g)	Total Amount (ng H-28548)	Percent
001F 002F 003F 004F 005F	73080 69600 69890 70760 73370	168 h 168 h 168 h 168 h 168 h	227.935 230.137 257.292 212.796 244.357	12.9 14.4 15.3 39.3 15.1	2940 3314 3937 8363 3690 Mean SD	4.02 4.76 5.63 11.8 5.03 6.25 3.16

Appendix F Material Balance Material Balance

		001M	002M	003M	004M	005M	Mean	SD
urine	6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h Subtotal	20.9 9.32 29.5 16.7 6.19 4.07 2.38 1.41 1.20 91.7	18.5 13.0 21.3 17.7 8.58 4.45 2.74 1.66 1.30 89.2	11.7 20.3 22.3 24.6 6.49 4.37 2.46 1.76 1.27 95.3	9.21 13.5 23.1 13.7 8.35 4.21 2.26 1.83 1.71 77.9	10.4 27.2 24.0 16.5 6.60 3.52 2.26 1.98 1.06 93.6	14.1 16.7 24.1 17.8 7.24 4.12 2.42 1.73 1.31 89.5	5.19 7.12 3.20 4.06 1.13 0.37 0.20 0.21 0.25 6.91
feces	6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h Subtotal	0.19 0.38 0.25 0.16 0.10 0.04 <loq <loq <loq< td=""><td>0.06 0.13 0.19 0.20 0.12 0.12 0.10 0.05 0.08</td><td>1.07 0.09 0.11 0.21 0.458 0.30 0.14 <loq <loq 2.38</loq </loq </td><td>0.15 0.15 0.34 1.36 0.21 0.36 0.15 0.399 0.40 3.51</td><td>0.08 0.50 0.60 0.50 0.08 0.073 0.047 <loq 0.05 1.92</loq </td><td>0.31 0.25 0.30 0.48 0.19 0.18 0.11 0.22 0.18 2.00</td><td>0.43 0.18 0.19 0.51 0.16 0.14 0.05 0.25 0.20</td></loq<></loq </loq 	0.06 0.13 0.19 0.20 0.12 0.12 0.10 0.05 0.08	1.07 0.09 0.11 0.21 0.458 0.30 0.14 <loq <loq 2.38</loq </loq 	0.15 0.15 0.34 1.36 0.21 0.36 0.15 0.399 0.40 3.51	0.08 0.50 0.60 0.50 0.08 0.073 0.047 <loq 0.05 1.92</loq 	0.31 0.25 0.30 0.48 0.19 0.18 0.11 0.22 0.18 2.00	0.43 0.18 0.19 0.51 0.16 0.14 0.05 0.25 0.20
cage wash	168 h	11.00	10.72	6.75	14.98	4.74	9.64	3.99
	Total	103.8	101.0	104.4	96.3	100.3	101.2	3.22
		001F	002F	003F	004F	005F	Mean	SD
urine	6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h Subtotal	12.4 24.0 26.8 18.11 5.15 4.03 1.55 0.79 0.61 93.4	19.3 19.1 21.5 17.5 5.16 3.05 1.72 2.05 1.25 90.7	22.7 26.4 21.6 12.8 7.53 2.49 1.82 0.74 0.49 96.7	18.5 17.7 18.9 13.8 4.87 3.24 2.00 1.46 1.04 81.5	13.0 23.4 21.7 20.3 9.34 4.13 1.63 1.03 0.90 95.4	17.2 22.1 22.1 16.5 6.41 3.39 1.74 1.22 0.86 91.5	4.41 3.60 2.88 3.11 1.96 0.69 0.17 0.55 0.31 6.04
feces feces feces feces feces feces feces feces feces	6 h 12 h 24 h 48 h 72 h 96 h 120 h 144 h 168 h Subtotal	0.28 0.07 0.17 0.49 0.09 0.14 <loq <loq <loq< td=""><td>1.16 0.06 0.32 0.19 0.43 0.09 0.302 0.260 0.060 2.87</td><td>0.08 0.07 0.08 0.77 0.10 <loq 0.05 <loq <loq 1.15</loq </loq </loq </td><td>0.42 0.66 0.24 0.72 0.28 0.10 0.16 0.17 0.064 2.80</td><td>0.33 0.37 0.16 0.35 0.28 <loq <loq <loq <loq< td=""><td>0.45 0.25 0.19 0.50 0.24 0.11 0.17 0.21 0.06 1.91</td><td>NA 0.27 0.09 0.24 0.14 0.03 0.12 0.07 0.00 0.85</td></loq<></loq </loq </loq </td></loq<></loq </loq 	1.16 0.06 0.32 0.19 0.43 0.09 0.302 0.260 0.060 2.87	0.08 0.07 0.08 0.77 0.10 <loq 0.05 <loq <loq 1.15</loq </loq </loq 	0.42 0.66 0.24 0.72 0.28 0.10 0.16 0.17 0.064 2.80	0.33 0.37 0.16 0.35 0.28 <loq <loq <loq <loq< td=""><td>0.45 0.25 0.19 0.50 0.24 0.11 0.17 0.21 0.06 1.91</td><td>NA 0.27 0.09 0.24 0.14 0.03 0.12 0.07 0.00 0.85</td></loq<></loq </loq </loq 	0.45 0.25 0.19 0.50 0.24 0.11 0.17 0.21 0.06 1.91	NA 0.27 0.09 0.24 0.14 0.03 0.12 0.07 0.00 0.85
cage wash	168 h	4.02	4.76	5.63	11.8	5.03	6.25	3.16

Appendix G Elimination Half-Life

Elimination Half-Life

OriginLab v7.0220, interpolation of mean urinary excretion data; interpolated data points every 3 hours from 0 to 168 hours (56 data points)

 $T_{1/2}$ Males: 21 hours $T_{1/2}$ Females: 18 hours

Bolded/underlined values (*) identify the elimination half-lives ($\geq 50\%$ of the administered dose) and associated cumulative percent of H-28548 in urine

Time, post-dose		H-28548 eliminated in urine
(hours)	Male	Female
0	-2.6	-4.9
3.05455	5.90182	6.35091
6.10909	14.40364	17.60182
9.16364	22.90545	28.85273
12.21818	31.23818	39.70182
15.27273	37.37273	45.32727
18.32727*	43.50727	50.95273*
21.38182*	49.64182*	56.57818
24.43636	55.22364	61.7
27.49091	57.48909	63.8
30.54545	59.75455	65.9
33.6	62.02	68
36.65455	64.28545	70.1
39.70909	66.55091	72.2
42.76364	68.81636	74.3
45.81818	71.08182	76.4
48.87273	72.96545	78.13273
51.92727	73.89455	78.94727
54.98182	74.82364	79.76182
58.03636	75.75273	80.57636
61.09091	76.68182	81.39091
64.14545	77.61091	82.20545
67.2	78.54	83.02
70.25455	79.46909	83.83455
73.30909	80.22364	84.48545
76.36364	80.74545	84.91818
79.41818	81.26727	85.35091
82.47273	81.78909	85.78364
85.52727	82.31091	86.21636
88.58182	82.83273	86.64909
91.63636	83.35455	87.08182
94.69091	83.87636	87.51455
97.74545	84.27455	87.83091
100.8	84.58	88.06
103.85455	84.88545	88.28909
106.90909	85.19091	88.51818
109.96364	85.49636	88.74727
113.01818	85.80182	88.97636
116.07273	86.10727	89.20545
119.12727	86.41273	89.43455
122.18182	86.65455	89.60909
125.23636	86.87091	89.76182
128.29091	87.08727	89.91455
131.34545	87.30364	90.06727
134.4	87.52	90.22
137.45455	87.73636	90.37273
140.50909	87.95273	90.52545
143.56364	88.16909	90.67818
146.61818	88.34182	90.78727
149.67273	88.50727	90.78727
152.72727	88.67273	90.99091
155.78182	88.83818	91.09273
158.83636	89.00364	91.19455

Time, post-dose	Cumulative percent of	H-28548 eliminated in urine
(hours)	Male	Female
161.89091	89.16909	91.29636
164.94545	89.33455	91.39818
168	89.5	91.5